Scheme I


The product distribution from the $3^{\prime}$ labeled RNA strand is compared to that from the $3^{\prime}$ labeled noncoding DNA strand of the Eco R 1 restriction fragment by using I as the nucleolytic agent (Figure 2). In both cases, the major cutting sites are clustered between positions 19-24. This interval of predominant cutting sites 2-3 nucleotides in both directions from the tethered 1,10phenanthroline can be due to the diffusibility of the oxidative species or the flexibility of the 1,10 -phenanthroline linked terminal deoxyadenosine that can be inferred from the RNase H hybridization studies. The kinetics of the cutting reaction are similar with both RNA and DNA. After incubation for 2 h at $37^{\circ} \mathrm{C}$, approximately $20 \%$ of the parent band is converted to one of the oligonucleotide products.

Primer extension assays were also used to monitor the reaction of I with RNA. ${ }^{13}$ These assays reflect phosphodiester bond scission as well as any oxidative damage that may block polymerization. Since the pattern of products obtained were similar to those using $3^{\prime}$ labeled RNA, there is no evidence for reaction that does not lead to strand scission. In previous studies of the DNase activity of 1,10 -phenanthroline-copper ion, no reaction without strand scission has ever been observed. ${ }^{14,15}$

The similarity in the digestion patterns suggests that the phosphodiester backbones of RNA and DNA are comparably reactive to the chemical nuclease activity of 1,10 -phenanthroline-copper. The extension of these findings to other oxidative nucleolytic activities, e.g., ferrous-EDTA and iron porphyrins, ${ }^{16-20}$ will require direct experimental tests in view of

[^0]bleomycin's inability to nick RNA.
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## Regio- and Stereoselective Reduction of N(21),N(22)-Bridged Porphyrin Hydroperchlorates to Stable 5H-Phlorins

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Hydroporphyrins with saturated meso carbon(s) are important in the redox chemistry and the biosynthesis of porphyrin. ${ }^{1}$ Phlorins, $5 \mathrm{H}, 22 \mathrm{H}$-dihydroporphyrins, are usually so air-sensitive that only a few have been fully characterized so far ${ }^{2}$ including those with the steric crowding of peripheral substituents which is relieved upon hybridization change from $\mathrm{sp}^{2}$ to $\mathrm{sp}^{3}$ of the meso carbon as shown in the Woodward's approach to chlorophyll $a .^{3}$

[^1]Scheme I ${ }^{a}$

$x^{-}$

${ }^{a} \mathrm{Q}: \mathrm{HC}=\mathrm{CH}(1,6), \mathrm{PhC}=\mathrm{CPh}(2,7,11-13),(o-\mathrm{Tol}) \mathrm{C}=\mathrm{C}(o-\mathrm{Tol})$ $(3,8), \mathrm{CH}_{2} \mathrm{CH}_{2}(4,9), \mathrm{C}=\mathrm{C}\left(\mathrm{C}_{6} \mathrm{H}_{4}-\mathrm{p}-\mathrm{Cl}\right)_{2}(5,10) ; \mathrm{X}: \mathrm{H}(\mathrm{D})(6-10)$, CN (11), $\mathrm{CH}_{2} \mathrm{COMe}$ (12), $\mathrm{CH}_{2} \mathrm{COPh}$ (13).

Table I. ${ }^{1} \mathrm{H}$ Chemical Shift ( $\delta$ ) and Spin-Lattice Relaxation Time ( $T_{1}$ ) of 5-Meso Methylene Protons of 5 H -Phlorins 6-8 ${ }^{\text {a }}$

| compd | $\mathrm{R}^{\text {b }}$ | endo-H |  | exo-H |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | $\delta$ | $T_{1}$ | $\delta$ | $T_{1}$ |
| 6 | H | 5.05 | 0.20 | 3.33 | 0.27 |
| 7 | $\mathrm{C}_{6} \mathrm{H}_{5}$ | 5.36 | 0.19 | 3.53 | 0.25 |
| 8 | $o-\mathrm{CH}_{3} \mathrm{C}_{6} \mathrm{H}_{4}$ | 5.42 | 0.14 | 4.15 | 0.15 |

${ }^{a} \delta$ and $T_{1}$ were measured in $\mathrm{CDCl}_{3}$. ${ }^{b}$ Substituent on the etheno bridge as shown in Figure 1.

This paper describes a new means to stabilize phlorin structure through the introduction of an $\mathrm{N}(21), \mathrm{N}(22)$-bridging group which exerts a strain on a porphyrin plane so that 5 -meso carbon favors a tetrahedral configuration.

Treatment of THF solution of $N(21), N(22)$-ethenooctaethylporphyrin hydroperchlorate ${ }^{4}$ (1) with $\mathrm{NaBH}_{4}$ (2.5-fold molar excess) under argon gave a blue compound which can be extracted into hexanes. Removal of hexanes afforded satisfactorily pure powders of $N(21), N(22)$-ethenooctaethyl- $5 H$-phlorin ${ }^{5}(6)$ in $73 \%$ yield. 6 was cleanly air-oxidized to 1 under acidic conditions. The visible spectrum of 6 is characteristic of a phlorin chromophore. ${ }^{6}$ ${ }^{1} \mathrm{H}$ NMR spectrum of 6 is indicative of the disappearance of a ring current effect and the presence of a symmetry plane which contains a $\mathrm{C}(5)-\mathrm{C}(15)$ axis and bisects the $\mathrm{N}(21), \mathrm{N}(22)$-bridge. $N(21), N(22)$-(1,2-Diphenyletheno)- and $N(21), N(22)$-(1,2-di( $o$ tolyl)etheno) octaethylporphyrin hydroperchlorate, ${ }^{4,7}$ (2) and (3), were analogously reduced to the corresponding $5 H$-phlorins, ${ }^{8} 7$ and 8 , in $84 \%$ and $45 \%$ yields, respectively. One of the two AB doublets due to the saturated meso methylene protons appeared at $3.3-4.2 \mathrm{ppm}$ and the other at $5.0-5.5 \mathrm{ppm}$ in the ${ }^{1} \mathrm{H}$ NMR spectra of 6-8 (see Table I). Irradiation of the methyl signal ( 1.52 ppm ) of the bridge $o$-tolyl groups of 8 resulted in $11 \% \mathrm{NOE}$ enhancement of only the higher field AB doublet ( 4.15 ppm ). A positive NOE effect was also observed between the doublet (5.77 ppm) due to the bridge ortho-phenyl protons of 7 and the higher field $A B$ doublet ( 3.53 ppm ). Therefore, the higher field $A B$

[^2]

Figure 1. A preferred boat form conformation of 6-8.
doublet is associated with the 5 -meso (not 15 -meso) methylene proton at the same side as the bridge (exo side), meaning that reduction takes place regioselectively at the 5 -meso position which is surrounded by the $N(21), N(22)$-bridge. This assignment of the AB doublets is consistent with the fact that spin-lattice relaxation time $\left(T_{1}\right)$ of the higher field doublet decreases in a larger quantity than that of the lower field doublet due to an increase in the number of the etheno bridge protons which can interact with the 5 -exo proton, as shown in Table I. Furthermore, the fact that $T_{1}$ of the endo proton is always shorter than that of the exo proton implies that the endo proton is in close proximity to the 3 - and 7 -ethyl group. This requires the seven-membered ring consisting of $N(21), C(4), C(5), C(6), N(22)$, and two etheno bridge carbons of the $5 H$-phlorins to take a boat form rather than a chair form (see Figure 1).
$N(21), N(22)$-Ethano- and $N(21), N(22)$-(bis $(p$-chlorophenyl)vinylideno)octaethylporphyrin hydroperchlorate, ${ }^{9,10}$ (4) and (5), were analogously reduced to the corresponding 5 H phlorins, ${ }^{11} 9$ and 10 , in $51 \%$ and $65 \%$ yields, respectively. Their spectral properties are quite similar to those of $6-8$. The higher field signal due to the 5 -exo proton disappeared completely, and the 5 -endo proton appeared as a singlet in the ${ }^{1} \mathrm{H}$ NMR spectra of the deuteriated 5 H -phlorins, $7 \mathrm{~d}, 9 \mathrm{~d}$, and 10 d , which were obtained by the reaction of 2,4 , and 5 with $\mathrm{NaBD}_{4}$. Therefore, a deuteride was incorporated exclusively at the 5 -exo position stereoselectively. Nucleophilic addition of natrium cyanide, kalium acetone enolate, and lithium acetophenone enolate to 2 occurred in preference to deprotonation, affording very stable 5 -substituted 5 H -phlorins, ${ }^{12} 11,12$, and 13 , in $74 \%, 84 \%$, and $81 \%$ yields, respectively. Since the chemical shifts of the saturated 5 -meso protons of 11-13 are very low ( $7.2-6.8 \mathrm{ppm}$ ), they are assignable to the endo side where a greater deshielding effect is exerted by the substituted pyrrole rings in a boat form. Thus, cyanide and enolates add to 2 regio- and stereoselectively in exactly the same manner as hydride.

The single-crystal X-ray analysis of $N(21), N(22)$-(1,2-diphenyletheno)tetraphenylporphin hydroperchlorate indicates that the etheno bridge forces the substituted pyrrole rings to be highly canted toward the endo side by $22.9^{\circ}$ and $20.6^{\circ}$ from the mean

[^3]

Figure 2. Exo-side attack of nucleophiles on 1-3 under the stereoelectronic control.
plane that contains four pyrrole nitrogens. ${ }^{7}$ As the tilt of the pyrrole rings causes a greater overlap of the p-orbital of $C(5)$ with that of $C(4)$ or $C(6)$ in the endo region than in the exo region, the anti-bonding $\pi$-orbital which interacts with nucleophiles should extend more to the exo side at $\mathrm{C}(5)$ (see Figure 2). This rationalizes the observed stereoselectivity, and a similar stereoelectronic effect should be expected for the $\mathrm{NAD}^{+}$model compounds ${ }^{13}$ which are closely related to $1-5$ in the sense that they are monocationic nitrogen heterocycles.

Finally, it should be emphasized that there are only a few examples of nucleophilic attack on the porphyrin system ${ }^{22,14}$ and that an $\mathrm{N}(21), \mathrm{N}(22)$-bridging group is removable in principle as has been demonstrated for the TPP analogue of $5 .{ }^{15}$ Thus, the present reaction is of importance not only as a new porphyrin redox system but also as a facile synthetic method for the meso-substitution of porphyrin by the use of ordinary carbanions.

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## Cupped- and Cappedophanes, Two New General Classes of Compounds with Molecular Cavities

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We describe here the first examples of two broad classes of molecules which should have potential as molecular hosts ${ }^{1}$ and other novel structural features. Their structures are based on an $m$-terphenyl framework in which the outer rings are orthogonal to the central ring. Compounds of type 1 may have any set of


[^4]Scheme I

$1 E$


Scheme II

atoms linking the $2,2^{\prime \prime}$ and $6,6^{\prime \prime}$ positions, thus forming a molecular bowl; as a consequence of the synthetic methodology, groups E other than hydrogen can also readily be incorporated, covalently bound to carbon $2^{\prime}$ in the middle of the bowl. Compounds of type 2, which resemble a canopied gondola in shape, have a capping unit linked to the "outer" terphenyl rings. ${ }^{2}$ If the connecting arms are long enough, these structures may include a passenger group E at position $2^{\prime}$.

Because of their shapes and cyclophane ${ }^{3}$ character, we refer to 1 and 2 as cupped- and cappedophanes, respectively. Several examples of each type, that can be synthesized in just a few steps, are described here.
The key intermediate 6 for the cuppedophanes and cappedophanes reported here was prepared as shown in Scheme I. The conversion of 3 to $\mathbf{4}$ involves tandem aryne reactions recently developed in our group, ${ }^{4}$ quenching allows the introduction of electrophiles (for example, deuterium) on the central ring at this stage.
Addition of a benzene solution of 6 and $m$-xylylenedithiol ${ }^{5}$ (2 equiv) under high dilution techniques ${ }^{6}$ to ethanolic KOH afforded tetrathia cuppedophane 7 in good yield. Oxidation gave the

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    (12) 11: ${ }^{1} \mathrm{H}$ NMR ( $\left.\mathrm{CDCl}_{3}, 270 \mathrm{MHz}\right) \mathrm{H}_{\text {mass }} 6.58$ (s) $(\times 2), 6.48$ (s), 7.06 (s); $\mathrm{CH}_{2} 2.9-2.5(\mathrm{~m}) ; \mathrm{CH}_{3} 1.39(\mathrm{t}), 1.36(\mathrm{t}), 1.05(\mathrm{t}), 0.92(\mathrm{t}) ; \mathrm{NH} 8.9(\mathrm{br})$; $\mathrm{H}_{\mathrm{p}} 6.76(\mathrm{t}) ; \mathrm{H}_{\mathrm{m}} 6.63(\mathrm{t}) ; \mathrm{H}_{0} 6.04(\mathrm{~d}) \mathrm{ppm} ; \mathrm{MS}, 737(\mathrm{M}) ; \mathrm{UV}-\mathrm{vis}\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right)$ $\lambda_{\text {max }} 382,620 \mathrm{~nm}$; Anal. satisfactory $\mathrm{C}, \mathrm{H}, \mathrm{N}$ for $\mathrm{C}_{51} \mathrm{H}_{55} \mathrm{~N}_{5}$. 12: ${ }^{i} \mathrm{H} \mathrm{NMR}$ $\left(\mathrm{C}_{6} \mathrm{D}_{6}, 270 \mathrm{MHz}\right) \mathrm{H}_{\text {meso }} 6.90(\mathrm{~s})(\times 2), 6.84(\mathrm{~s}), 6.87(\mathrm{t}) ; \mathrm{CH}_{2} \mathbf{3 . 4 - 2 . 5}(\mathrm{~m})$; $\mathrm{CH}_{3} 1.41(\mathrm{t}), 1.39(\mathrm{t}), 0.99(\mathrm{t})(\times 2) ; \mathrm{NH} 9.5(\mathrm{br}) ; \mathrm{H}_{\mathrm{m} . \mathrm{p}} 6.57(\mathrm{~m}) ; \mathrm{H}_{\mathrm{o}} 6.16$ (m); $\mathrm{COCH}_{2} 2.78$ (d); $\mathrm{COCH}_{3} 1.59$ (s) ppm; MS, 769 (M+1); UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) \lambda_{\max } 393,638 \mathrm{~nm}$; Anal. satisfactory $\mathrm{C}, \mathrm{H}, \mathrm{N}$ for $\mathrm{C}_{53} \mathrm{H}_{60} \mathrm{~N}_{4} \mathrm{O}$. 13: ${ }^{1} \mathrm{H}$ NMR ( $\left.\mathrm{C}_{6} \mathrm{D}_{6}, 270 \mathrm{MHz}\right) \mathrm{H}_{\text {meso }} 6.91$ (s) $(\times 2), 6.86$ (s), 7.21 (t); $\mathrm{CH}_{2}$ $3.4-2.5(\mathrm{~m}) ; \mathrm{CH}_{3} 1.45(\mathrm{t}), 1.40(\mathrm{t}), 0.98(\mathrm{t})(\times 2) ; \mathrm{NH} 9.5(\mathrm{br})$; bridge $\mathrm{H}_{\mathrm{p}}$ 6.54 (t); bridge $\mathrm{H}_{\mathrm{m}} 6.47$ (t); bridge $\mathrm{H}_{\mathrm{o}} 6.19$ (d); $\mathrm{COCH}_{2} 3.49$ (d); meso $\mathrm{H}_{0}$ 7.74 (d); meso $\mathrm{H}_{\mathrm{p}} 6.90$ (t); meso $\mathrm{H}_{\mathrm{m}} 6.71$ (t) ppm; UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right) \lambda_{\text {max }} 394$, 637 nm ; Anal. satisfactory $\mathrm{C}, \mathrm{H}, \mathrm{N}$ for $\mathrm{C}_{58} \mathrm{H}_{62} \mathrm{~N}_{4} \mathrm{O}$.

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